

# Integrated Model of the Eye/Optic Nerve Head Biomechanical Environment

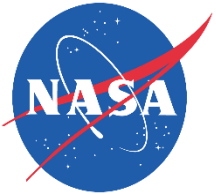
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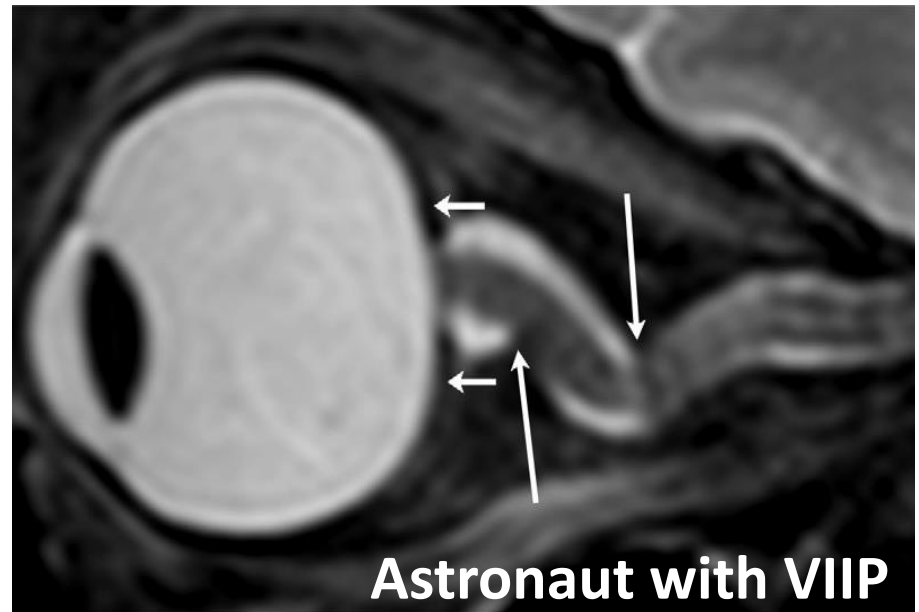
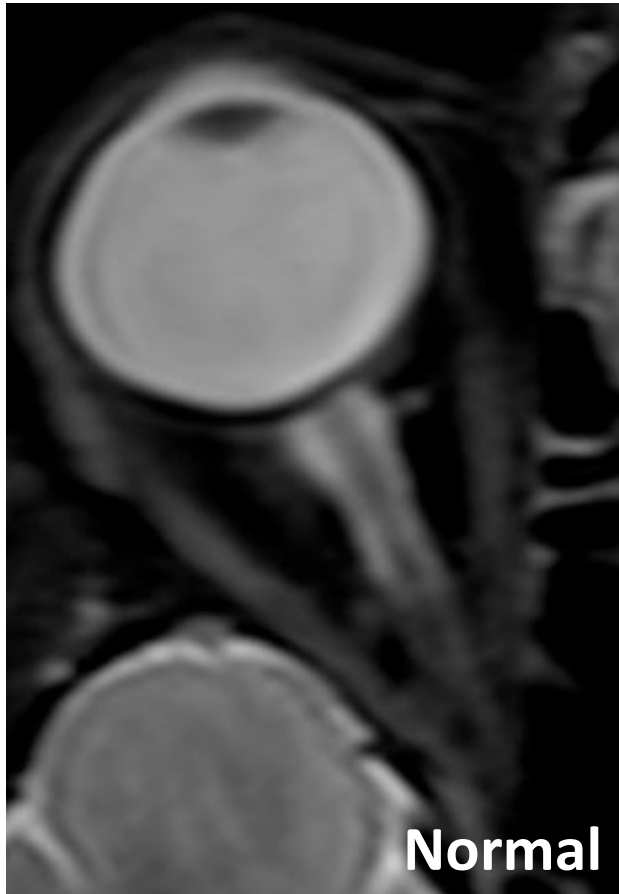
**Wallace H. Coulter** Department of  
**Biomedical Engineering**  
at Georgia Tech and Emory University



**EMORY**  
UNIVERSITY

# Structural Changes in the Posterior Eye

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Kramer et al. Radiology, 2012.

# Hypothesis

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Increased CSF pressure, transmitted to the RB-SAS, drives remodeling of connective tissues in the posterior eye and optic nerve sheath

Eventually leads to the vision disturbances characteristic of VIIP

# Goal

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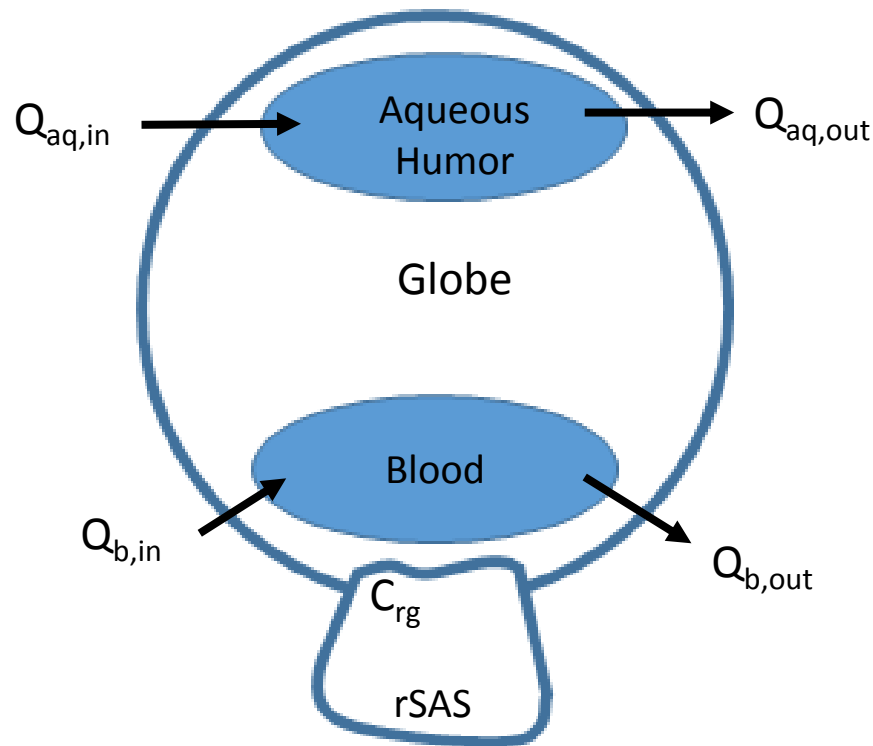
Develop an integrated model approach to understand how environmental conditions impact deformation of tissues of the posterior eye and optic nerve sheath

Key tools: Numerical and finite element modeling

# Numerical Model

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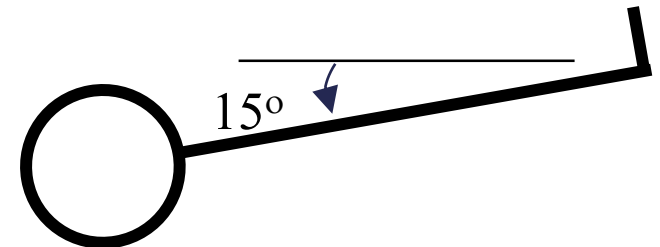
# Lumped Parameter Eye Model



## Model features:

- Four-compartment model
  - Anterior Chamber
  - Blood compartment (cardiovascular model)
  - Globe
  - Retrobulbar subarachnoid space (rSAS)

## Simulating Head Down Tilt (HDT)

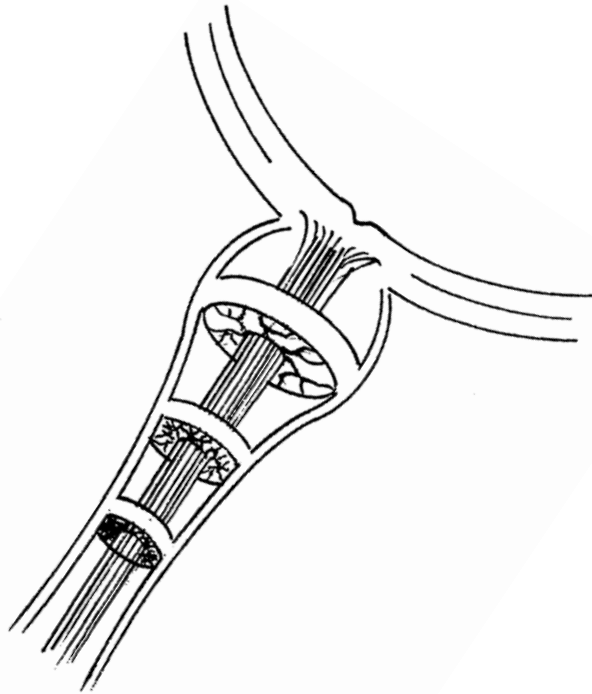


# Finite Element Model

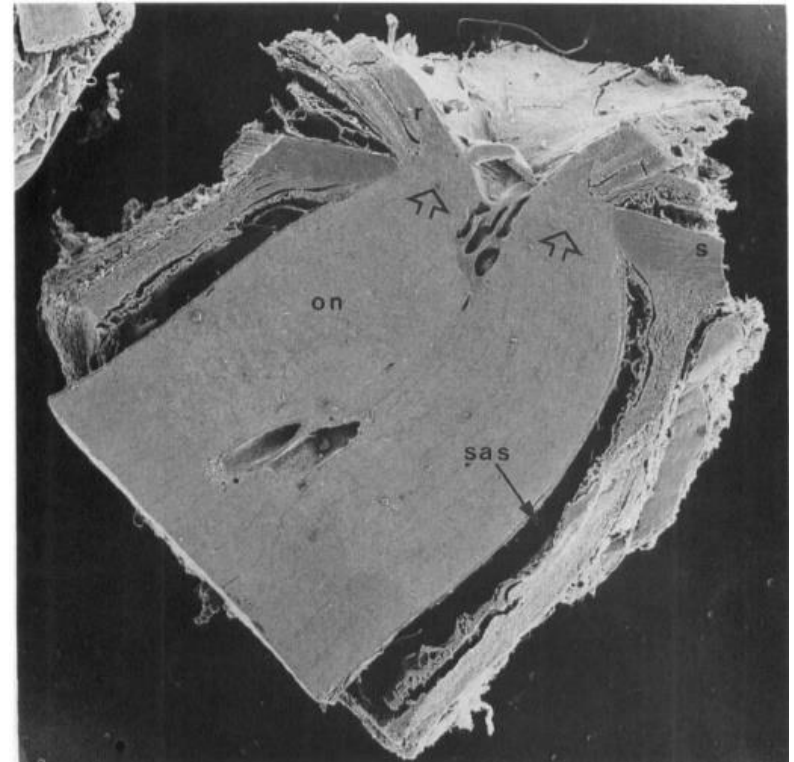
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# Finite Element Geometry

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Hansen et al. Acta Ophthalmologica, 2011.

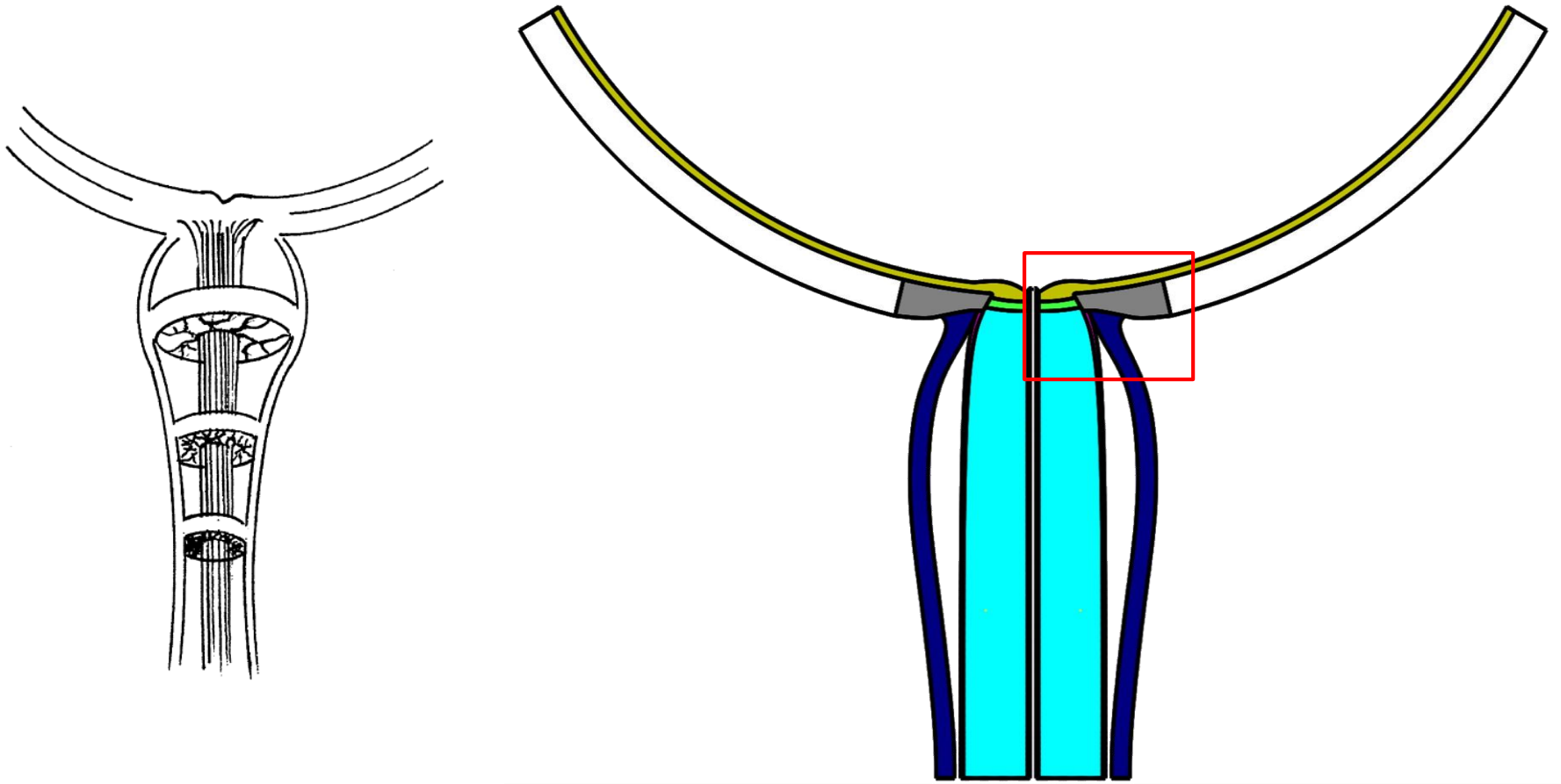


Adopted from Ekington et al. 1990

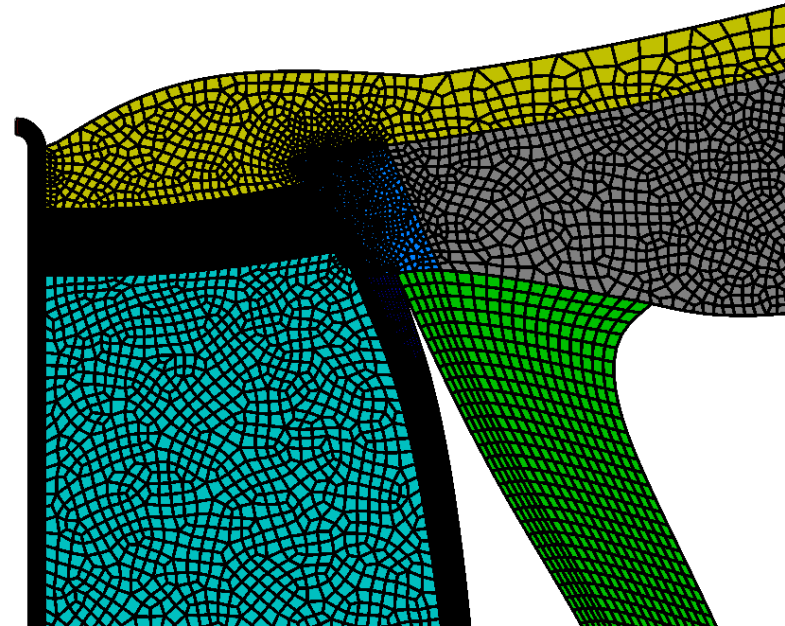
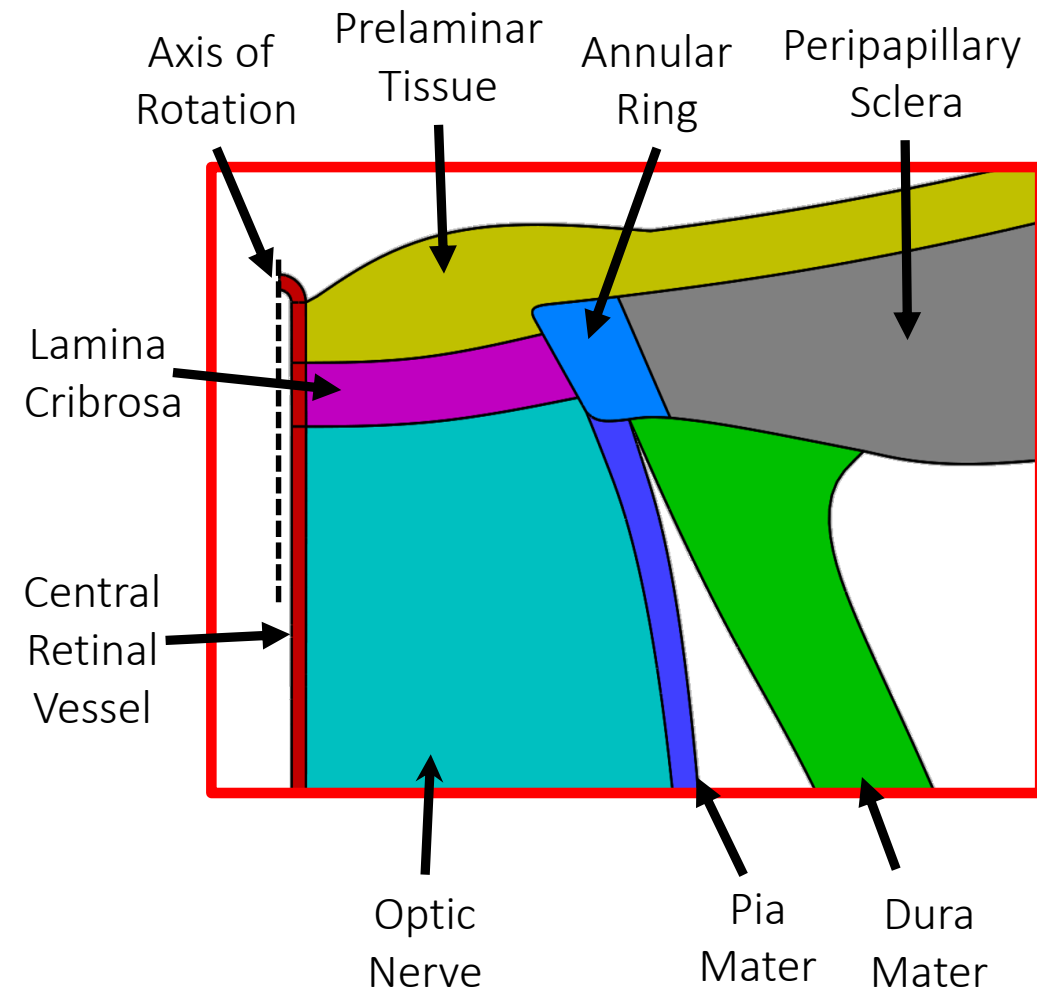


# Model Overview

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# Finite element model



# Tissue Constitutive Models

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- Mooney-Rivlin plus von Mises Distributed Fibers
  - Proposed by Girard and Ethier for the the sclera
  - Implemented into FEBio V2 by Gouget and Girard for thin tissues

$$\Psi = F_1(\tilde{I}_1, \tilde{I}_2) + \int_{\theta_p - \frac{\pi}{2}}^{\theta_p + \frac{\pi}{2}} P(\theta) F_2(\tilde{\lambda}[\theta]) d\theta + \frac{K}{2} [\ln(J)]^2$$

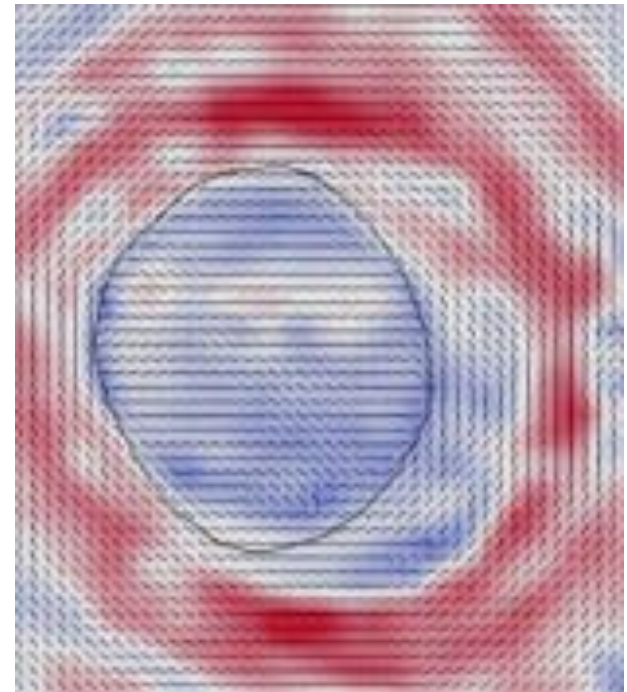
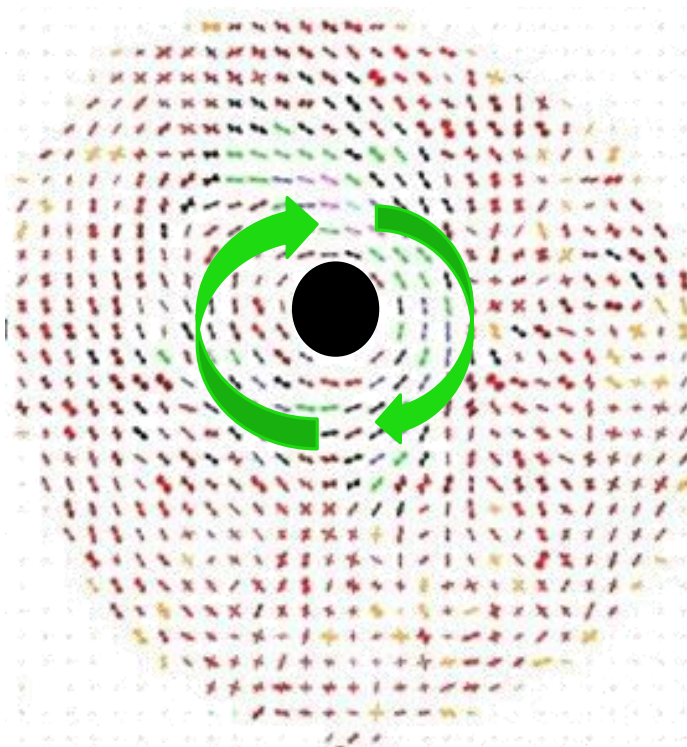
- $F_1$  represents ground substance (neo-Hookean):  $F_1 = c_1(\tilde{I}_1 - 3)$
- $F_2$  represents collagen fibers
  - Collagen fibers are loaded within their non-linear region

$$\tilde{\lambda} \frac{\partial F_2}{\partial \tilde{\lambda}} = 0, \tilde{\lambda} \leq 1$$
$$\tilde{\lambda} \frac{\partial F_2}{\partial \tilde{\lambda}} = c_3(e^{c_4(\tilde{\lambda}-1)} - 1), 1 < \tilde{\lambda} \leq \lambda_m$$

# Collagen Orientation in the Sclera

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- Sclera: collagen fibers treated as transversely isotropic
- Peripapillary sclera: moderately aligned collagen fibers
- Annular ring: highly aligned collagen fibers



~ Pijanka et al. 2012 & Zhang et al. 2015

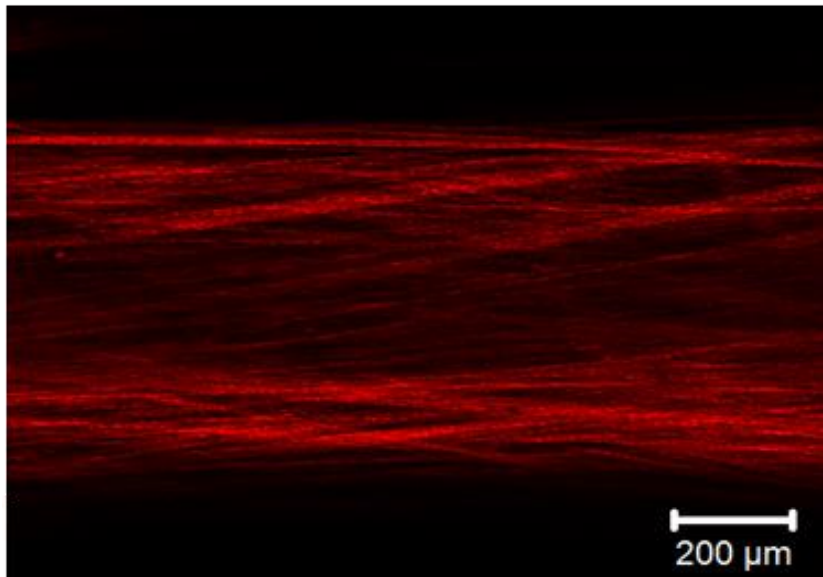
# Collagen Orientation in the ONS

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Pia mater and dura mater: fibers were modeled as transversely isotropic

~Raspanti et al. 1992 Noort et al. 1980 & Raykin et al. 2015

## Dura Mater

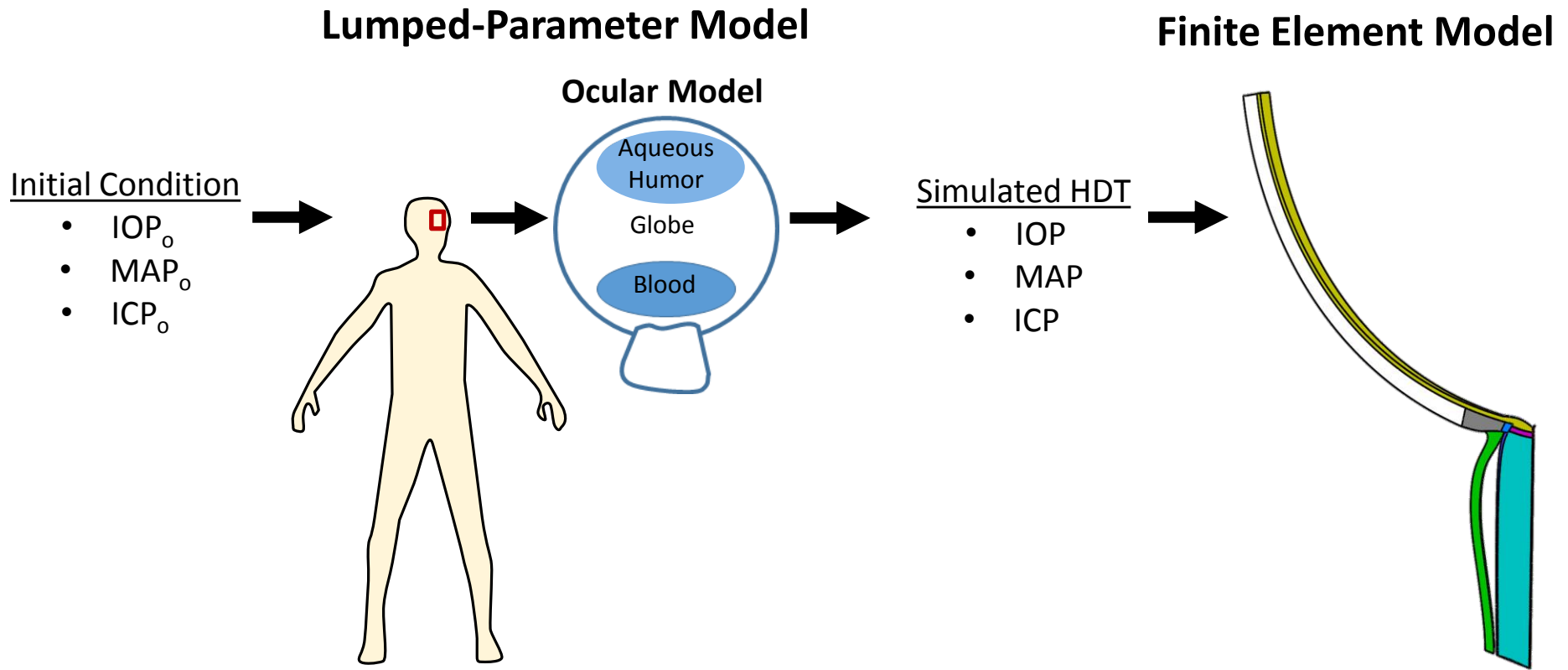


# Tissue Material Properties

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	Mooney-Rivlin solid with embedded collagen fibers
Material Inputs	Ground Stiffness ( $c_1$ ) Collagen Stiffness ( $c_3$ & $c_4$ )
Tissue Properties	Distribution of embedded collagen fibers

# Integration Overview



# Outcome measures

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- Strain (fractional tissue elongation) in all tissue regions
  - Strain is a tensor and can be decomposed into 3 primary components
    - First principal strain (stretch)
    - Second principal strain
    - Third principal strain (compression)
- Why do we care about strain?
  - Cells are mechanosensitive and alter their phenotype in response to mechanical strain

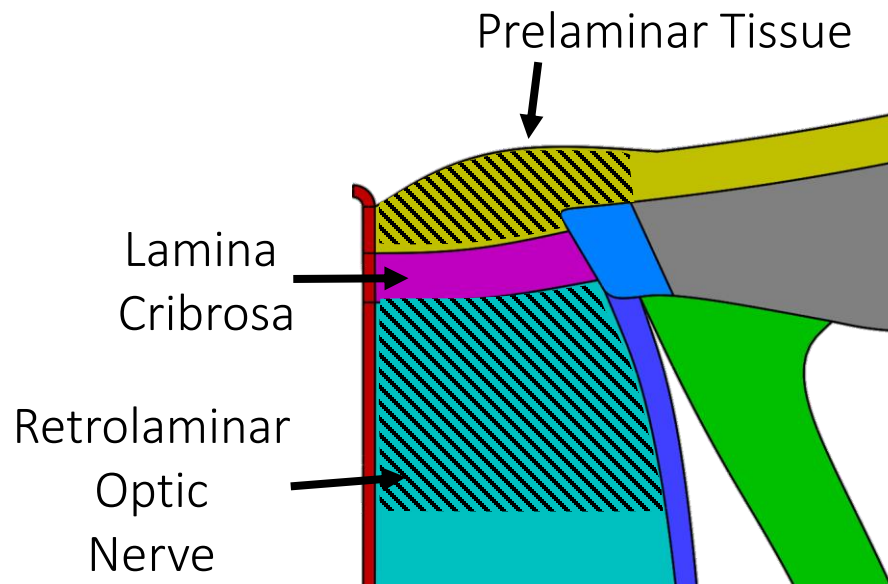


# Finite Element Model

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Primary outcome measures: peak tensile and compressive strains in the prelaminar tissue, lamina cribrosa and retrolaminar optic nerve

## Regions of Interest:

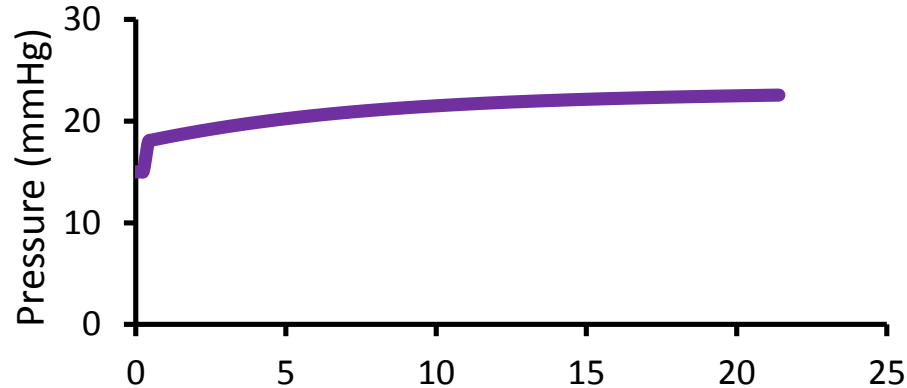


# Results

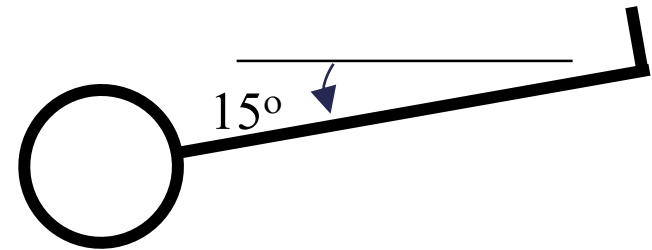
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# Pressures from Eye Model

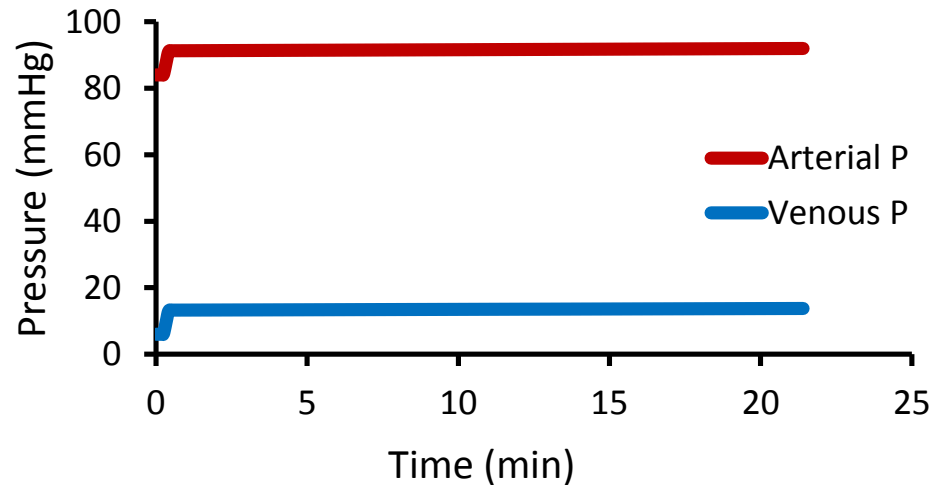
Intraocular Pressure



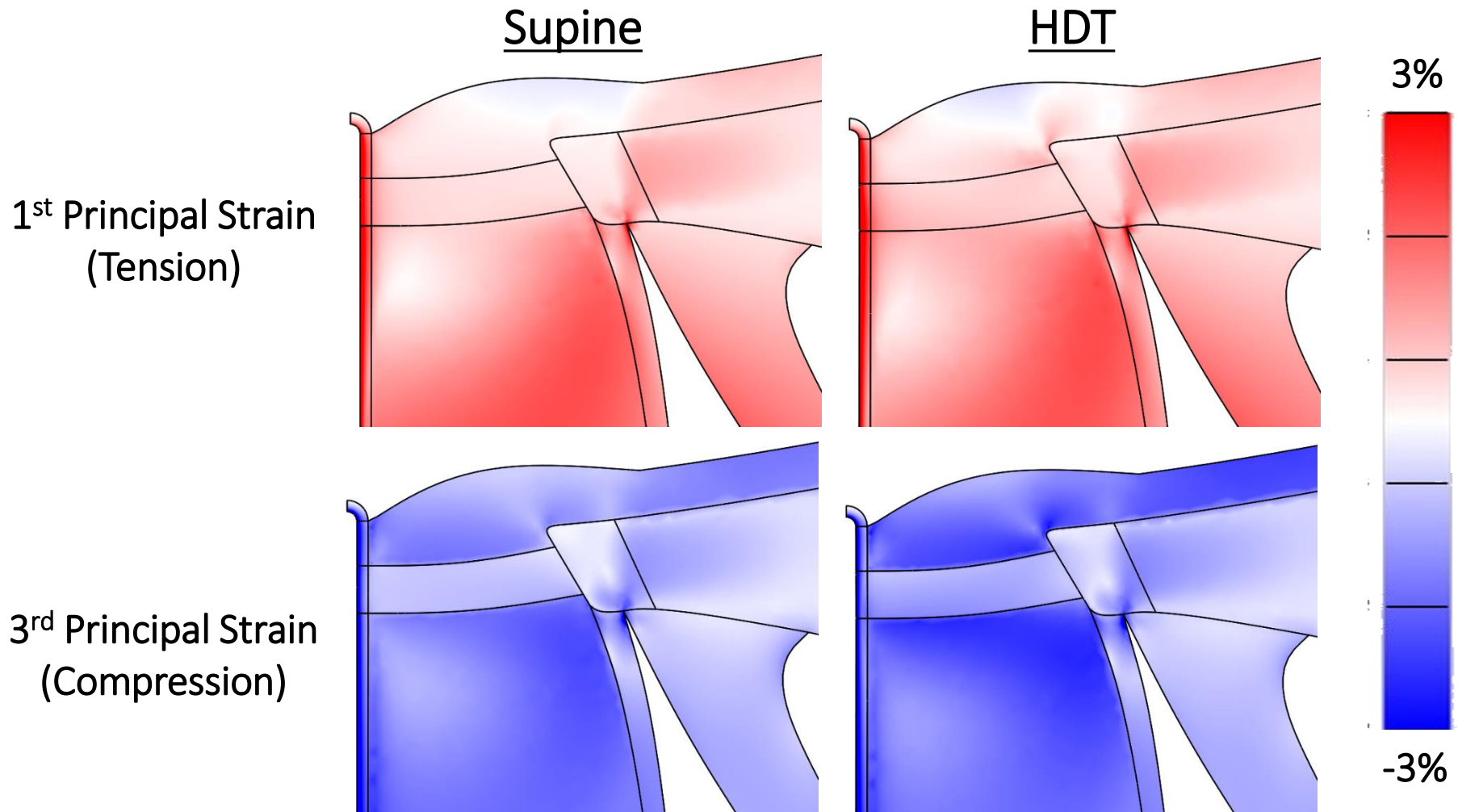
HDT



Blood Pressure



# Principal Strain Magnitudes



# HDT on ONH Deformation

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		<u>Supine</u>	<u>HDT</u>
Lamina Cribrosa	Tension	0.60%	0.93%
	Compression	-0.98%	-1.51%
Retrolaminar Optic Nerve	Tension	1.16%	1.39%
	Compression	-1.64%	-2.44%
Prelaminar Neural Tissue	Tension	0.77%	1.25%
	Compression	-1.75%	-2.69%

# Summary

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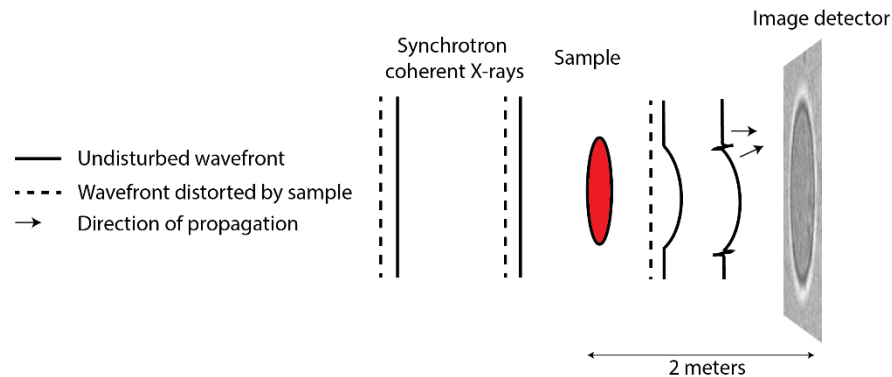
- Our integrated model approach predicts an increase in strains at the ONH after HDT
- These strains, if persistent, may induce a phenotypical change in ONH cells
- Future experimental work should examine how strains initiate a remodeling response in the optic nerve and optic nerve sheath

# Experimental Effects of ICP on the Optic Nerve

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# Experiment Objective

- Measure strain in the optic nerve due to elevations in ICP
  - Refraction of the X-ray by the sample
  - Tissues can be intact and untreated (no contrast agent required)



- However, for resolution of our small complex tissues requires X-rays with high spatial coherence

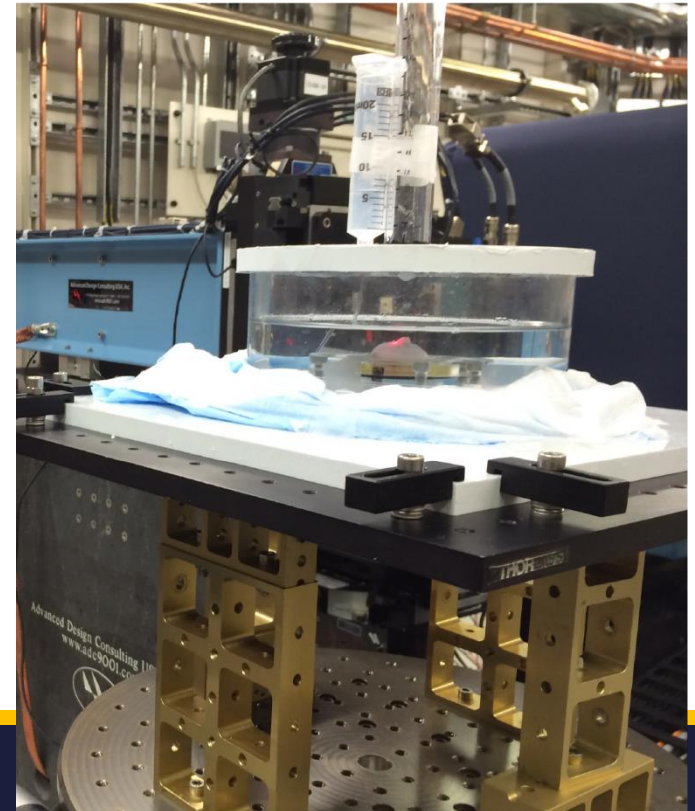
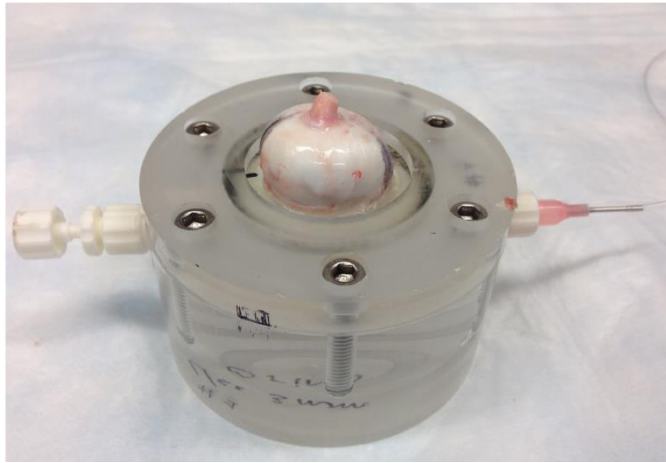




# Experimental Design

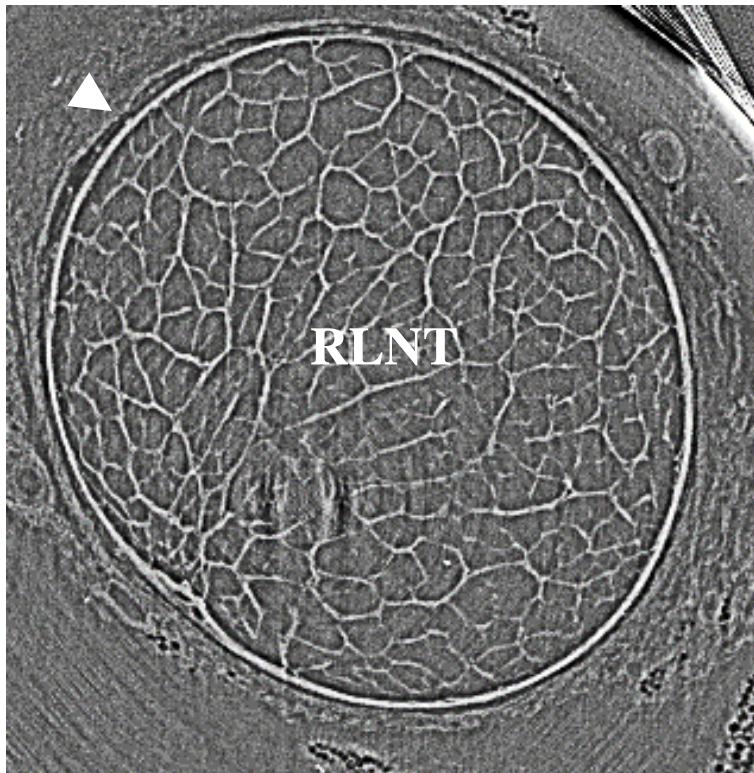
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- 3 porcine eyes
- Micro-CT scans were acquired at an ICP = 4, 10, 20 & 30 mmHg
- IOP was kept constant at 15 mmHg

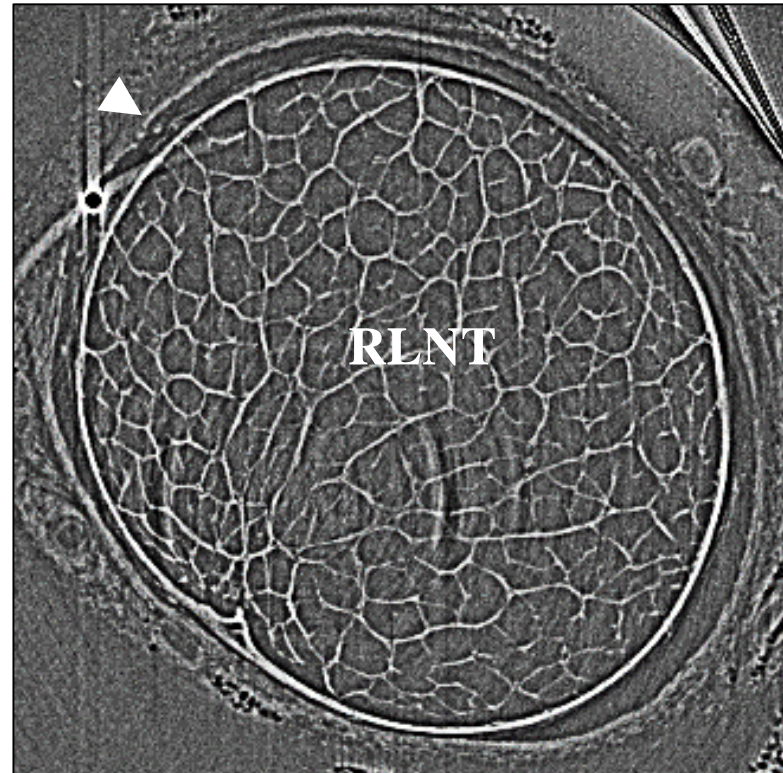


# Phase-contrast micro-CT

Non-uniform expansion of the dura mater



**CSFp = 4 mmHg**



**CSFp = 30 mmHg**

# Optic Nerve Deformation

ICP

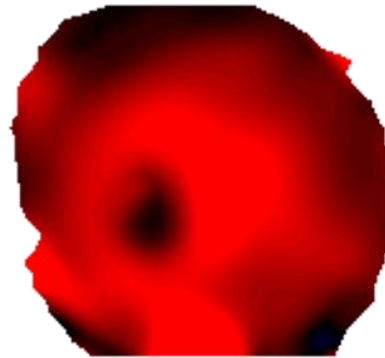
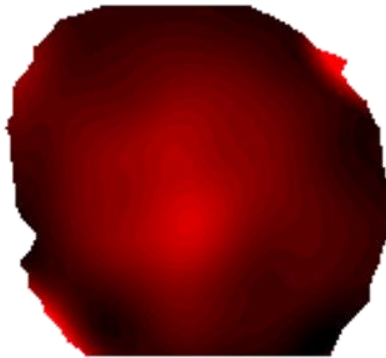
CSFp:

10 mmHg

20 mmHg

30 mmHg

1<sup>st</sup>  
Principal  
Strain

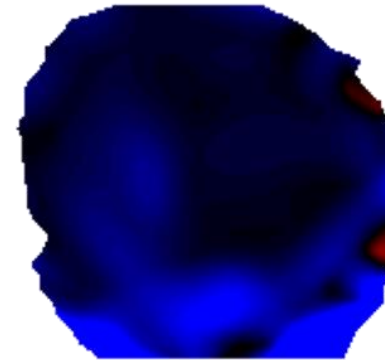
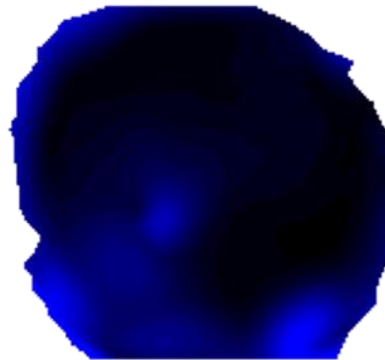


10%



-10%

3<sup>rd</sup>  
Principal  
Strain



# Experimental Work Summary

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- Increased ICP directly elevated strains in the optic nerve
- Our experimental results agree with earlier finite element model work
  - The magnitude of strain was higher in experimental results

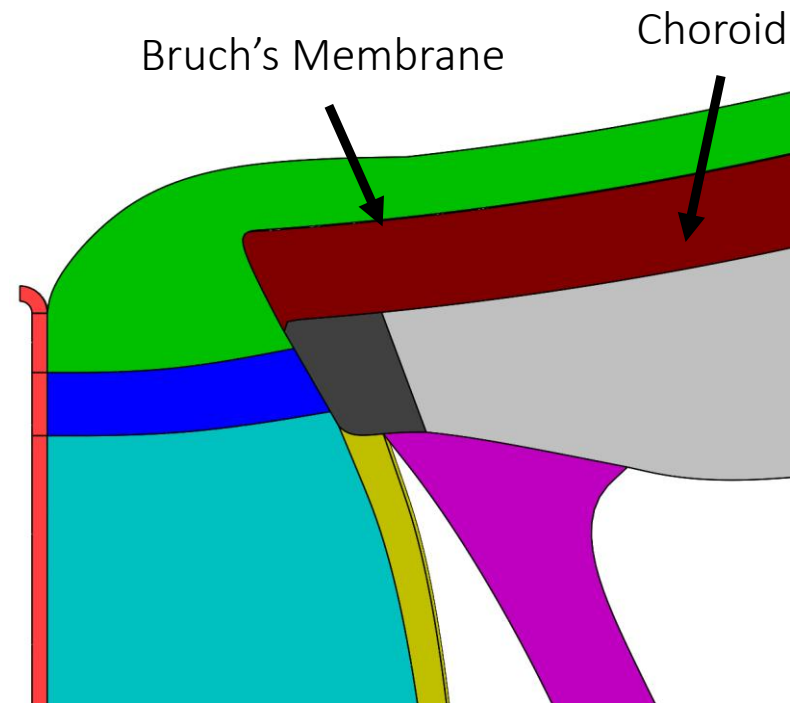
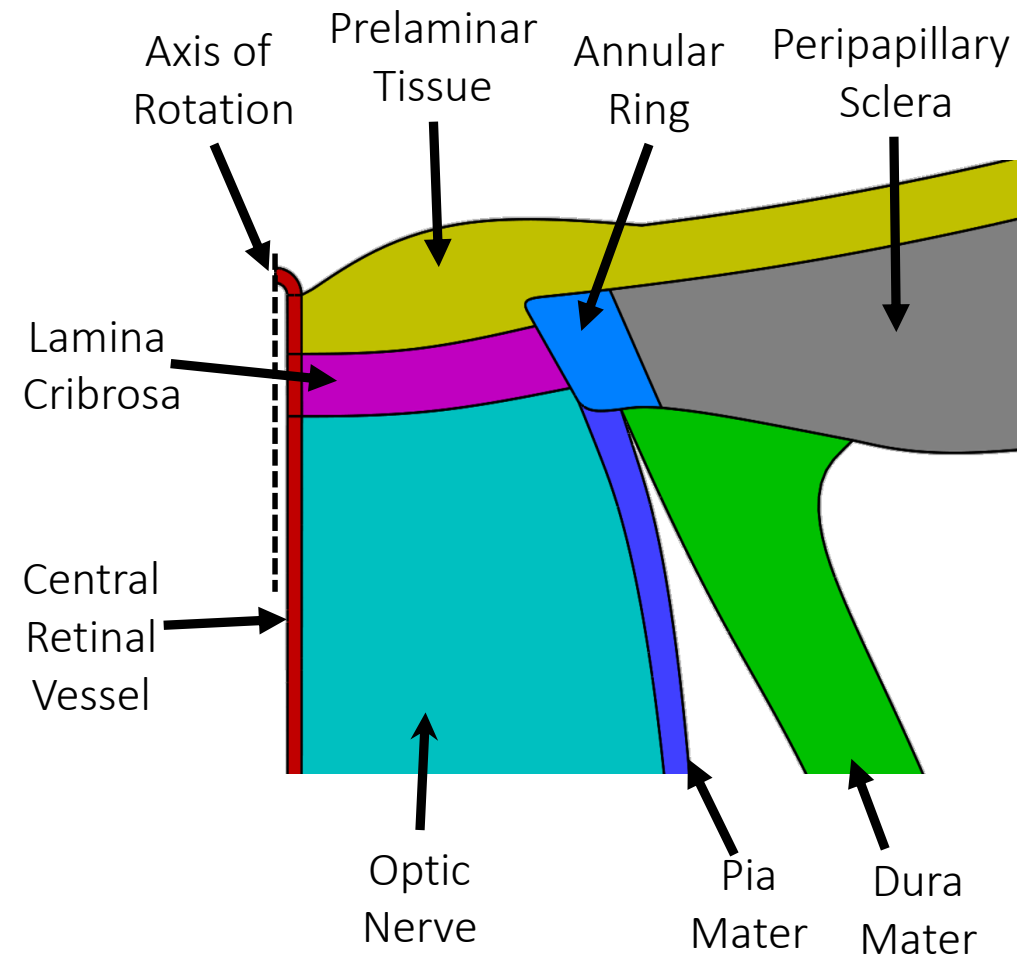
# Ongoing Work

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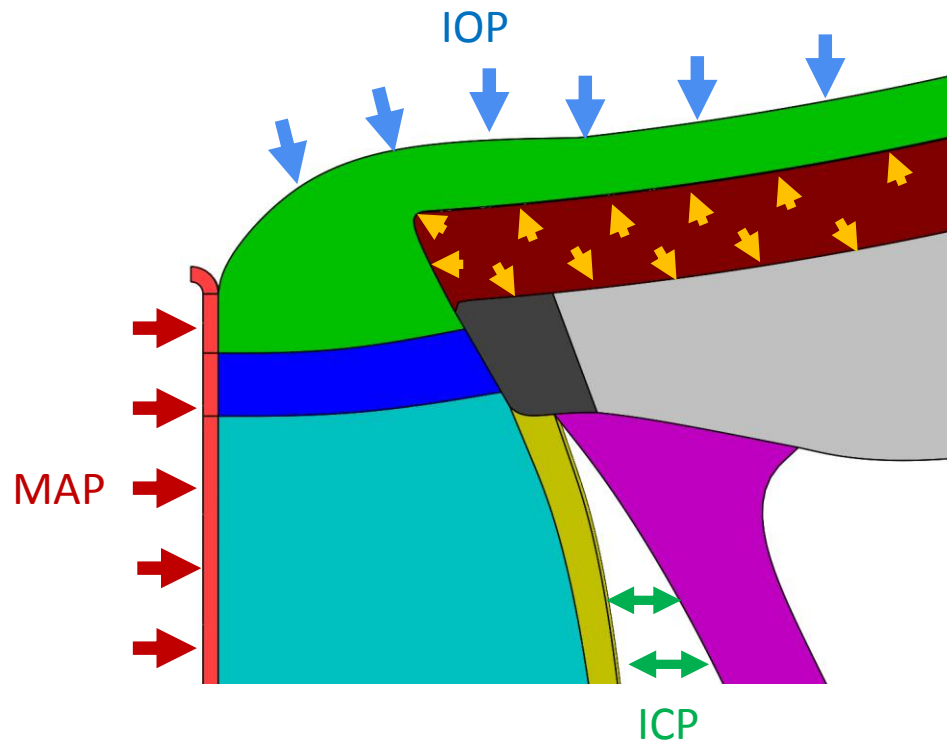
# Additional FE Work

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# Simulate Choroidal Swelling

- Choroid modeled as a solid mixture to allow swelling
  - Linear-Elastic material ( $E = 0.3 \text{ MPa}$ )
  - Apply uniform swelling (5  $\mu\text{L}$ ) to simulate volume change during cardiac cycle

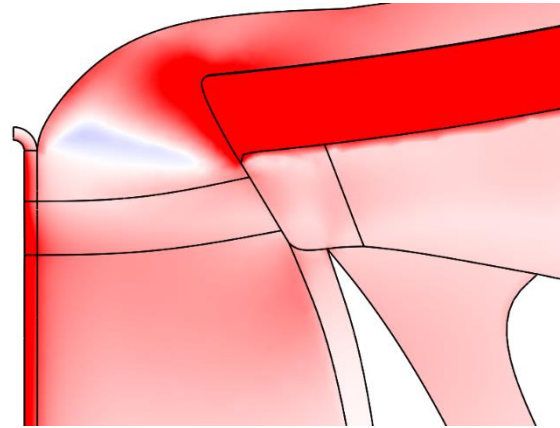
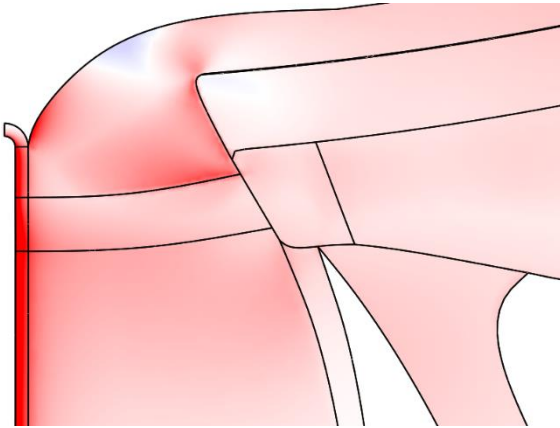


# Impact of Choroidal Swelling

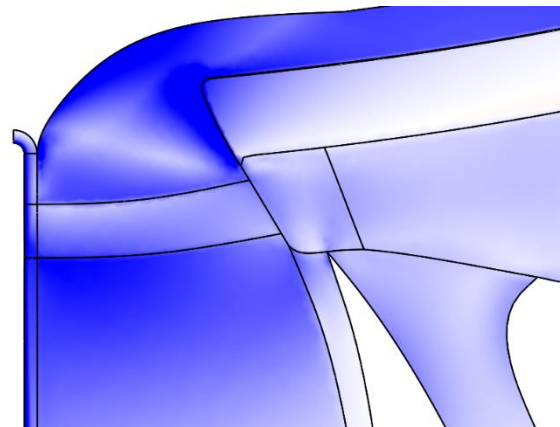
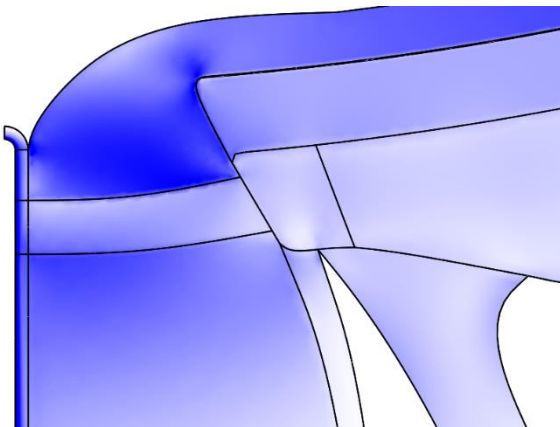
IOP = 15 mmHg  
Choroid Swelling = 0 uL

IOP = 15 mmHg  
Choroid Swelling = 5 uL

1<sup>st</sup> Principal  
Strain



3<sup>rd</sup> Principal  
Strain



2%



-2%

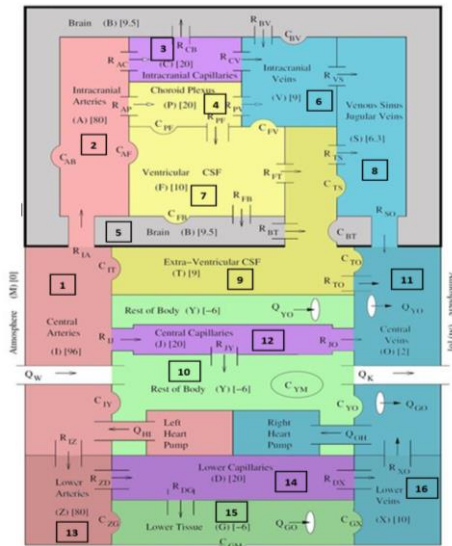


# Advanced Model Integration

## Latin Hypercube Sampling Inputs

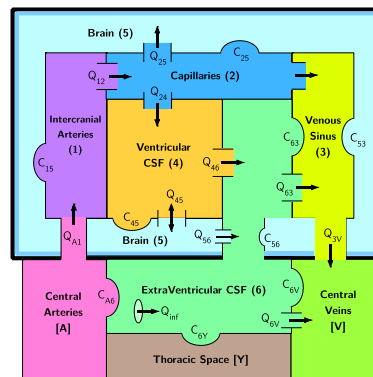
	Cardiovascular	Central Nervous	Eye Model	FEM
Run 1	$w_1 \dots w_{42}$	$x_1 \dots x_{17}$	$y_1 \dots y_3$	$z_1 \dots z_{20}$
Run 2	$w_1 \dots w_{42}$	$x_1 \dots x_{17}$	$y_1 \dots y_3$	$z_1 \dots z_{20}$
Run N	$w_1 \dots w_{42}$	$x_1 \dots x_{17}$	$y_1 \dots y_3$	$z_1 \dots z_{20}$

## 16 Compartment Cardiovascular System

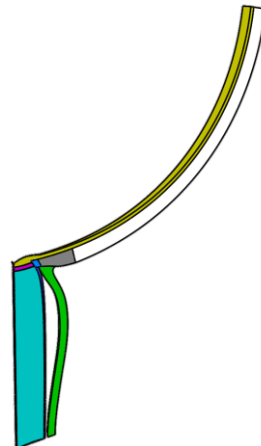
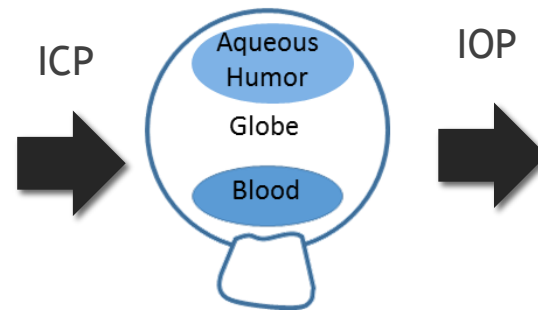


## 6 Compartment Central Nervous System

Blood Pressure



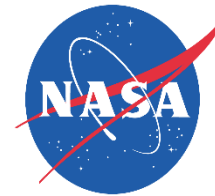
## 4 Compartment Eye



# Acknowledgements

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- NASA support, grant NNX13AP91G
- Drs. DeVon Griffin and Beth Lewandowski



# BME at Georgia Tech/Emory

